

Treatment of Acute Lymphoblastic Leukemia

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Abstract

Acute lymphoblastic leukemia (ALL) affects both children and adults, with prevalence between the ages of 2 and 5 years. Serial clinical trials have resulted in steady improvement in the outcome of patients with ALL. Most children and over a third of adult patients are cured with widely available treatment approaches based on the use of risk-directed multiagent chemotherapy regimens and diligent supportive care. Ongoing research is now aiming at reducing long term treatment sequelae in children and younger adults, and at improving the outcome of adults and some subgroups of children with poor prognosis. This review tracks six decades of progress in the therapy of ALL, summarizes the rationale of contemporary ALL therapy, and addresses remaining challenges.

Introduction

Steady progress in the development of treatment strategies for acute lymphoblastic leukemia (ALL) started in the 1950s, when complete remissions were achieved using single chemotherapeutic agents.^{1,2} Significant improvement in remission duration was achieved in pediatric trials combining and cycling these agents, with the addition of central nervous system (CNS) prophylaxis.³ Further progress was accomplished with serial clinical trials using outcome predictors to stratify therapy.⁴⁻¹² Drugs developed over 30 years ago, such as mercaptopurine, methotrexate, vincristine, corticosteroids, anthracyclines, and asparaginase, still constitute the backbone of contemporary ALL protocols resulting in long-term, event-free survival rates exceeding 80% in children, but seldom exceeding 40% in adult patients.⁴ With improved understanding of the immunology and molecular pathways involved in ALL, current risk classification typically include age, leukocyte count at diagnosis, blast cell immunophenotype and genotype, as well as early treatment response. The favorable hyperdiploidy >50 (more than 50 chromosomes) and TEL-AML1 gene fusion (with expression of ETV6-CBFA2) are present in about 50% of childhood ALL, and rarely seen in adults. On the other hand, the unfavorable Philadelphia chromosome (BCR-ABL) is present in about 25% of adults, but is not common in children.¹³⁻¹⁴ Early response to therapy, as measured by minimal residual disease (MRD) evaluation, has played an increasingly important role in risk stratification of ALL. Early and vigorous assessment of the risk of relapse in individual patients help improve outcome of patients with high-risk leukemia while minimizing long term sequelae and enhancing the quality of life in patients at low risk of relapse.⁴ Historically, adult ALL trials included more intensive alkylating agents based chemotherapy. Recently, adult investigators are exploring pediatric protocols in the young adult population, and innovative approaches in high risk subgroups.

Principles of treatment

Accurate assessment of relapse hazard is an integral part of ALL therapy. The prognostic impact of age, and to a lesser extent, leukocyte count can be explained

partly by their association with specific genetic abnormalities. For example, the poor prognosis of infants is associated with MLL rearrangement (detected in 70% to 80% of patients in this age group), and the overall favorable outcome of patients aged 1 to 9 years is related to the preponderance of cases with hyperdiploidy >50 or TEL-AML1 fusion.^{4,14} However, primary genetic features do not entirely account for treatment outcome. While up to 15% of patients with hyperdiploidy >50 or TEL-AML1 fusion suffer recurrences of their leukemia, a substantial proportion of the patients with the t(9;22) and BCR-ABL fusion who are 1 to 9 years old and have low leukocyte counts at diagnosis may be cured with intensive chemotherapy alone.¹⁵ Among patients with MLL-AF4 fusion, infants and adults have a worse prognosis than children.¹⁶⁻¹⁸ Interindividual variability in the pharmacokinetics and pharmacodynamics of many antileukemic agents might partially explain the heterogeneity in treatment response among patients with specific genetic abnormalities and the difference in outcome by age group. Multiple genetic polymorphisms have been associated with relapse risk, acute toxicity, and late effects.¹⁹⁻²⁴ The prime example of optimizing therapy based on germline genetic status is the use of polymorphisms of thiopurine methyltransferase (TPMT), an enzyme that catalyzes the methylation of thiopurines such as mercaptopurine and thioguanine, to guide treatment.²³ Some drugs affect outcome when administered concomitantly with ALL therapy. Drugs that induce cytochrome P450 enzymes (e.g. phenobarbital and phenytoin), significantly increase the systemic clearance of several antileukemic agents and may adversely affect treatment outcome. On the other hand, drugs that inhibit cytochrome P450 enzymes (e.g. azole antifungal and macrolide antibiotics), potentiate the effects of vincristine, anthracyclines and etoposide resulting in increased toxicity.

Response to therapy is determined by several factors including the leukemic cell biologic features, the pharmacogenetics of the patient, the treatment regimens administered, and compliance to therapy. The degree of reduction of the leukemic cell clone early during remission induction therapy has greater prognostic strength than any other individual biological or host related feature.²⁵ Assessing MRD by flow-cytometric detection of aberrant immunophenotypes or analysis by polymerase chain reaction (PCR) of clonal antigen-receptor gene rearrangements, provides a level of sensitivity and specificity that cannot be attained by traditional morphological assessment of treatment response.²⁵ There is strong concordance between the assessment of MRD by flow cytometry and by PCR methods. Over 95% of patients can be followed by flow cytometry which is a simple and rapid method. PCR method could be reserved for the few patients whose leukemic cells lack a suitable immunophenotype.

Phases of therapy

With the exception of mature B-cell ALL cases, which are treated with short-term intensive chemotherapy (including high-dose methotrexate, cytarabine, and cyclophosphamide), therapy for ALL typically consists of a brief remission-induction phase followed by intensification (or consolidation) therapy to eliminate residual disease, and then prolonged continuation treatment to maintain remission.

All patients also require treatment directed to the CNS early in the clinical course to prevent relapse due to leukemic cells sequestered in this site.⁴

Contemporary pediatric protocols stratify therapy based on risk groups defined by age, leukocyte count, immunophenotype, leukemic genotype, and response to early remission induction therapy.⁵⁻¹² The standard or lower risk category includes patients between 1 and 10 years of age with an initial leukocyte count less than $50 \times 10^9/L$, and the remaining patients are considered higher risk. Additional features used by some investigators to stratify patients as lower risk include hyperdiploidy > 50 (DNA index > 1.16) and trisomy of chromosomes 4 and 10. Conversely, other characteristics, such as T-cell phenotype, adverse cytogenetic translocations [t(9;22) and t(4;11)], overt CNS leukemia at diagnosis, and slow early response to induction chemotherapy, have been used to stratify patients as high risk.

Remission induction

Remission induction therapy aims at eradicating leukemic cell burden and restoring normal hematopoiesis. This treatment phase typically lasts 4 to 6 weeks, and includes the administration of a glucocorticoid (prednisone or dexamethasone), vincristine, and at least a third drug (asparaginase or anthracycline, or both). A two-drug remission induction regimen of weekly vincristine and daily prednisone results in remission in 80% to 90% of children with ALL.^{26,27} Addition of a third agent, such as asparaginase or an anthracycline, increases the remission rate to approximately 95%.^{28,29} In addition to improving remission rates, intensified induction regimen also prolong remission duration.^{29,30} A three-drug induction regimen appears sufficient for most standard-risk cases, provided they receive intensified postremission therapy.³¹ The benefit in long-term survival of using 4 or more drugs during induction is widely accepted in higher risk patients but less clear in lower risk patients.³² Addition of a tyrosine kinase inhibitor has greatly improved the remission induction rate, duration of disease-free survival and quality of life of patients with Philadelphia positive (Ph+) ALL.³³⁻³⁵

Based on reports of more potent in-vitro antileukemic activity and better CNS penetration,³⁶⁻³⁹ dexamethasone has replaced prednisone in some induction and many continuation regimens.^{12,40-42} However, the biologically equivalent doses between dexamethasone and prednisone are not known, and one study suggested that prednisone can yield results comparable to dexamethasone, provided higher dose is used (i.e. 60 mg/m²/day).⁴³ Dexamethasone given at higher dose was associated with increased incidence of hyperglycemia, hypertension, myopathy, bony morbidity, severe behavioral changes and infectious complications.⁴⁴ As with glucocorticoids, the pharmacodynamics of asparaginase differ by formulation.⁴⁵ The native *E. coli* asparaginase has been the most commonly used preparation. Polyethylene glycol-conjugated asparaginase, a long-acting and less allergenic form, is progressively replacing the native product and is being increasingly administered intravenously instead of intramuscularly.⁴⁶ Asparaginase derived from

Erwinia chrysanthemi, has a short half-life and its use is currently limited to patients who are allergic to the *E. coli* formulations. The dose schedule for asparaginase should take into account the variability in the pharmacokinetic profile and potency among the different preparations.

The rapidity of response to induction therapy, as measured by clearance of peripheral and bone marrow blasts, is an important predictor of outcome,^{4,5} although intensification of postinduction therapy can improve the adverse prognosis of slow early responders.⁴⁷ With modern chemotherapy and supportive care, 97% to 99% of children can be expected to attain complete morphological remission (i.e. $< 5\%$ blasts in bone marrow) at the end of remission induction; those who do not have poor outcome.^{48,49} Hence, most investigators offer these patients the option of allogeneic hematopoietic stem cell transplantation at the end of extended induction treatment.⁵⁰ We and others have found that patients with 1% blasts identified by MRD studies had an outcome as poor as those with induction failure and the patients may also be candidates for allogeneic transplantation following intensification therapy to reduce MRD prior to transplant.^{51,52}

Consolidation (Intensification)

Following remission induction, consolidation (or intensification) is given to eradicate drug-resistant residual leukemic cells. Therapy is tailored to the leukemia subtype and risk-group. Intensifying asparaginase therapy during the early phase of treatment improved results of Dana Farber Cancer Institute (DFCI) studies,⁵³

adding doxorubicin to asparaginase favorably influenced the outcome of high-risk patients, particularly those with T-cell disease.^{54,55} Significant improvement was also reported in the outcome of patients receiving early intensification consisting of intermediate-dose or high-dose antimetabolite therapy.⁵⁵⁻⁵⁸

Delayed intensification, pioneered by the Berlin-Frankfurt-Münster (BFM) consortium, consists of using drugs similar to those used in remission induction therapy after a three months period of a less intensive, interim maintenance chemotherapy.⁵ The Children's Cancer Group (CCG) confirmed the efficacy of delayed re-induction therapy in low-risk cases,⁵⁹ and showed that double-delayed intensification with a second re-induction at week 32 of treatment, improved outcome in patients with intermediate-risk disease.⁶⁰ An augmented intensification regimen consisting of the administration of additional doses of vincristine and asparaginase during the myelosuppression period following delayed intensification, and sequential escalating-dose parental methotrexate followed by asparaginase (Capizzi methotrexate), improved the outcome of high-risk patients whose disease had responded slowly to initial multiagent induction therapy.⁴⁷

Continuation (Maintenance)

Continuation or maintenance phase consists of 2 to 2.5 years of low intensity metronomic chemotherapy designed to eradicate any residual leukemic cell burden. Weekly low-dose methotrexate and daily oral mercaptopurine form the backbone of most continuation regimens. Adjusting chemotherapy doses to maintain neutrophil counts between 0.5 and $1.5 \times 10^9/L$ has been associated with a better clinical outcome.^{4,61} Overzealous use of mercaptopurine, to the extent that neutropenia necessitates chemotherapy interruption, reduces overall dose intensity and is counterproductive.⁶² It is generally recommended to give mercaptopurine at bedtime to patients with an empty stomach,⁶³ and to avoid taken it together with milk or milk products which contain xanthine oxidase, an enzyme that can degrade the drug.⁶⁴ About 10% of the population inherit one wild-type gene encoding TPMT and one nonfunctional variant allele, resulting in intermediate enzyme activity, while 1 in 300 people inherits two nonfunctional variant alleles and are completely deficient of this inactivating enzyme.^{65,66} Patients with heterozygous and especially homozygous deficiency of TPMT are at high risk of severe myelosuppression. Identification of these patients allows to selectively guide reductions in mercaptopurine dosage without modifying the dose of methotrexate.^{23,67} Patients with TPMT deficiency are also at greater risk of developing therapy-related acute myeloid leukemia and radiation-induced brain tumors, in the context of intensive thiopurine therapy.^{23,68-70} Substituting thioguanine for mercaptopurine during continuation therapy was associated with a high incidence of profound thrombocytopenia and hepatic veno-occlusive disease.⁷¹⁻⁷³ Thioguanine use has therefore been limited to short pulses administered during consolidation therapy in some trials, while mercaptopurine is selected for prolonged administration.

Many groups add regular pulses of vincristine and corticosteroids to this regimen although the benefit of these pulses in the context of contemporary therapy has not been established.⁷⁴ The optimal duration of therapy remains unknown. Attempts to shorten therapy duration from 24 months to 12 or 18 months have resulted in a significant increase in relapses.⁷⁵ Many studies extend treatment for boys to 3 years because of their generally poorer outcome compared with girls,^{76,77} although the benefit of this approach remains to be demonstrated. Several studies showed no advantage to prolonging treatment beyond 3 years.^{78,79}

CNS directed therapy

The importance of therapy directed to the CNS was first demonstrated by investigators at St. Jude Children's Research Hospital in the 1960s, when the incidence of CNS leukemia as an initial site of relapse became progressively more common as more effective chemotherapeutic regimens resulted in longer duration of hematologic remissions. This was attributed to the CNS acting as a pharmacologic sanctuary, poorly penetrated by conventional doses of systematically administered chemotherapeutic agents.^{3,80,81} Radiation therapy was the first modality successfully used to prevent CNS relapse.⁸² The effectiveness of 2400 cGy cranial radiation as preventive therapy was offset by substantial late effects in long-term survivors, including learning disabilities, multiple endocrinopathy, and an increased risk of second malignancies. Subsequent trials demonstrated that, in the context of intensive systemic and intrathecal therapy, cranial irradiation can

be reduced 5,83 or even omitted altogether. 5,8,83,84

Because cranial irradiation can cause many acute and late complications (eg, second cancers, neurocognitive deficits, endocrine disorders and growth impairment), it has been largely replaced by intensive intrathecal treatment and systemic chemotherapy. Prophylactic cranial irradiation (12-18 Gy) given to patients with ALL who have an increased risk of CNS relapse restricts CNS relapse to 3-8% of patients. Patients with high-risk genetic features, T-cell immunophenotype, large leukemic burden, poor response to remission induction treatment, and leukemic cells in the cerebrospinal fluid (CSF) even from iatrogenic introduction from a traumatic lumbar puncture at diagnosis, are at increased risk of CNS relapse and require more intense CNS-directed therapy.^{85,86} Special care should be taken to minimize traumatic lumbar punctures, to deliver intrathecal therapy optimally, and to intensify systemic and intrathecal therapy in high-risk cases.⁸⁷ Studies have successfully used triple intrathecal therapy or intrathecal methotrexate alone.⁷² Systematically administered agents including high dose methotrexate,⁸⁸⁻⁹⁰ dexamethasone,³⁶ and asparaginase⁹¹ may contribute to prevention of CNS relapse.

Allogeneic hematopoietic stem-cell transplantation

Comparisons between allogeneic hematopoietic stem-cell transplantation and intensive chemotherapy have yielded inconsistent results due to the small numbers of patients studied and differences in case selection criteria.^{92,93} Allogeneic transplantation during initial complete remission may improve outcome of patients with poor response to initial induction chemotherapy, t(9;22), early hematologic relapse, or T-cell ALL with poor early response or hematologic relapse.^{15,35,94} The benefit of allogeneic hematopoietic stem-cell transplantation in infants with t(4;11) ALL remains controversial.^{17,95-97} Matched unrelated-donor or cord blood transplantation has yielded outcomes comparable to those obtained with matched related-donor transplantation, and should be considered reasonable alternatives if a matched donor is not available.^{98,99} Autologous transplantation has failed to improve outcome in ALL.⁹² With improving prospects for effective targeted therapy, the need for allogeneic transplantation should be continuously re-evaluated.

Challenging age groups

As cure rates approach 90% in children aged 1 to 9 year old, the following age groups still present challenges that need to be addressed with more innovative approaches.

Infants

> Infant

ALL comprises about 2% of total ALL cases (4% of childhood ALL). Whereas the outcome of the 15-20% infants with MLL germline ALL is comparable to that of older children with ALL, those with very young age (< 6 months), high initial leukocyte count (WBC > 300 x 10⁹/L), MLL rearrangement, and a poor early response to therapy have a dismal prognosis with less than 20% survival rates.^{17,95,100,101} The use of hematopoietic stem-cell transplantation in infants is controversial. Studies suggesting that the use of hematopoietic stem-cell transplantation contributed to a favorable outcome in infant ALL did not have a control arm in which patients only received chemotherapy and the data were not corrected for waiting time to hematopoietic stem-cell transplantation.^{97,98} Moreover, in one of these studies total body irradiation was used and led to substantial late effects in infants.⁹⁸ Data from a large retrospective intergroup analysis did not show differences between infant MLL rearranged cases who did or did not receive hematopoietic stem-cell transplantation.¹⁷

> Adolescents and young adults

Older adolescents and young adults (AYA) 16-21 years receive treatment from either pediatric or adult oncologists depending on referral pattern. Overall, the number of patients in this age range comprise a relatively small percentage of either pediatric or adult ALL trial populations, and they are often analyzed together with patients 10-15 years old in pediatric series, or those patients 20-30 years and older in adult clinical trials. Several retrospective analyses have demonstrated significantly better survival for AYA patients treated on pediatric cooperative group studies (event free survival 60-65%) compared with survival of patients from the same age group

who were treated on adult cooperative group trials (event free survival 30-40%). Pediatric protocols generally include more intensive use of nonmyelosuppressive agents (glucocorticoids, asparaginase, and vincristine), earlier and more intense CNS directed therapy, and more prolonged maintenance. Differences in adherence to protocol therapy among pediatric or adult medical oncologists and the patients they treat may also contribute to the discrepancy in survival. To understand the actual basis for this difference in outcome, several investigators and consortia are using common regimens to treat patients aged 1-50 years.

> Older adults

Despite improvements in the achievement of complete remission and progress in the supportive care of adults with ALL, the majority of patients eventually relapse, and the overall survival is only 30-40%. The Philadelphia chromosome (Ph+) resulting in the BCR-ABL fusion gene is the most common cytogenetic abnormality in adults with ALL, and is detected in approximately 50% of patients with B-precursor cell ALL who are over 60 years old. Elderly patients cannot tolerate intensive treatment, and trials including older adults have provisions for dose reductions in this age group. The incorporation of molecularly targeted therapy using the ABL tyrosine kinase inhibitor, imatinib mesylate, has begun to change the therapeutic landscape and outcome.³³ Ongoing trials are incorporating newer kinase inhibitors to overcome resistance. Addition of rituximab to the hyper-CVAD regimen appear to improve outcome in CD20+ patients, compared to CD20+ ALL on hyper-CVAD alone.¹⁰²

Relapse

Therapeutic options for refractory ALL are limited. Most relapses occur during treatment or within the first 2 years after its completion, although relapses have been reported as late as 10 years after initial ALL diagnosis.¹⁰³ The most common site of relapse is the bone marrow. Relapse in extramedullary sites, such as the CNS and testes, has decreased to less than 5% and 2% respectively.¹⁰⁴ Leukemia relapse occasionally occurs at other sites. Patients presenting with an isolated extramedullary relapse often have MRD in the bone marrow.¹⁰⁵ Patients with isolated bone marrow relapse generally fare worse than those with combined bone marrow and extramedullary relapse.¹⁰⁵ Factors indicating an especially poor prognosis are short initial remission and T-cell immunophenotype. Other adverse factors include t(9;22). The presence of minimal residual disease at the end of second remission induction is also a strong adverse prognostic indicator.^{106,107} Salvage regimens are mostly based on different combinations of the same agents used in frontline therapy, and are associated with significant morbidity and dismal long term survival rates in most cases. Patients with early or multiple relapses and heavy prior chemotherapy exposure, have an expected median survival of 9 to 10 weeks even when multiagent chemotherapy is used. While chemotherapy may secure a prolonged second remission in children with ALL who experience late relapse (defined as more than 6 months after cessation of therapy), allogeneic hematopoietic stem cell transplantation is the treatment of choice for patients who experience hematologic relapse during therapy or shortly thereafter and for those with T-cell ALL. Patients with late-onset isolated CNS relapse who had not received cranial irradiation as initial CNS-directed therapy have a very high remission retrieval rate, with long-term prognosis approaching that of newly diagnosed patients in those who had a long initial remission before the CNS event.^{92,108,109}

Future directions

Current therapy for patients with ALL has become increasingly dependent upon patient and disease-specific characteristics. Expanding the application of pharmacogenomics, a science which aims to define the genetic determinants of drug effects, will allow further individualized therapy in the future. While optimizing the use of old drugs continues through serial studies, new formulations of existing agents are being tested to improve the efficacy and reduce the toxicity of the parent compounds. Such modifications include improving drug transport and delivery, or altering the molecular structure to improve the therapeutic index. Ongoing trials are studying the benefit of the two novel nucleoside analogs, clofarabine and nelarabine, in high risk ALL and T-cell ALL respectively. In addition to refining leukemia classification, studies of global gene expression help identify potential molecular targets for therapy. It remains to be determined whether the success in targeting BCR-ABL with tyrosine kinase inhibitors will translate to other

pathways including NOTCH and FLT3. The challenge is to combine our current knowledge with technology to design effective risk-targeted therapies based on biological features of leukemic cells, host genetics, and early response to therapy. The dramatic increase that has occurred in the cure rate for children with ALL will be difficult to replicate in older patients without considerable additional research. In order to raise the survival rate of adolescents and adults with ALL, researchers will need a more thorough understanding of the biology of this form of leukemia, including the role that genes play in therapies.

References

1. Farber S, Diamond LK, Mercer RD, et al: Temporary remissions in acute leukemia in children produced by folic acid antagonist, 4-aminopteroyl-glutamic acid (aminopterin). *N Engl J Med* 1948; 238:787-793.
2. Frei E III, Holland JF, Schneiderman MA, et al: A comparative study of two regimens of combination chemotherapy in acute leukemia. *Blood* 1958;13:1126-1148.
3. Aur RA, Simone J, Hustu HO, et al: Central nervous system therapy and combination chemotherapy of childhood lymphocytic leukemia. *Blood* 1971;37:272-281.
4. Pui CH, Evans WE: Treatment of acute lymphoblastic leukemia. *N Engl J Med* 2006;354:166-178.
5. Schrappe M, Reiter A, Zimmermann M, et al: Long-term results of four consecutive trials in childhood ALL performed by the ALL-BFM study group from 1981 to 1995. *Berlin-Frankfurt-Munster. Leukemia* 2000;14:2205-2222.
6. Gaynon PS, Trigg ME, Heerema, NA, et al: Children's Cancer Group trials in childhood acute lymphoblastic leukemia: 1983-1995. *Leukemia* 2000;14:2223-2233.
7. Harms DO, Janka-Schaub, GE on behalf of the COALL Study Group. Co-operative study group for childhood acute lymphoblastic leukemia (COALL): long-term follow-up of trials 82, 85, 89 and 92. *Leukemia* 2000;14:2234-2239.
8. Vilmer E, Suciú S, Ferster S, et al: Long-term results of three randomized trials (58831, 58832, 58881) in childhood acute lymphoblastic leukemia: a CCG-EORTC report. *Leukemia* 2000;14:2257-2266.
9. Maloney KW, Shuster JJ, Murphy S, et al: Long-term results of treatment studies for childhood acute lymphoblastic leukemia: Pediatric Oncology Group studies from 1986-1994. *Leukemia* 2000;14:2276-2285.
10. Pui CH, Boyett JM, Rivera GK, et al: Long-term results of Total Therapy studies 11, 12 and 13A for childhood acute lymphoblastic leukemia at St. Jude Children's Research Hospital. *Leukemia* 2000; 14:2286-2294.
11. Tsuchida M, Ikuta K, Hanada R, et al: Long-term follow-up of childhood acute lymphoblastic leukemia in Tokyo Children's Cancer Study Group 1981-1995. *Leukemia* 2000;14: 2295-2306.
12. Silverman LB, Declerck L, Gelber RD, et al: Results of Dana-Farber Cancer Institute Consortium protocols for children with newly diagnosed acute lymphoblastic leukemia (1981-1995). *Leukemia* 2000; 14: 2247-2256.
13. Larson S, Stock Wendy: Progress in the treatment of adults with acute lymphoblastic leukemia. *Current Opinion in Hematology* 2008;15:400-407.
14. Pui CH, Relling MV, Downing JR. Acute Lymphoblastic Leukemia. *N Engl J Med* 2004;350:1535-1548.
15. Arico M, Valsecchi MG, Camitta B, et al: Outcome of treatment in children with Philadelphia chromosome-positive acute lymphoblastic leukemia. *N Engl J Med* 2000;342:998-1006.
16. Mancini M, Scappaticci D, Cimino G, et al : A comprehensive genetic classification of adult acute lymphoblastic leukemia (ALL): analysis of the GIMEMA 0496 protocol. *Blood* 2005;105:3434-3441.
17. Pui CH, Gaynon PS, Boyett JM, et al: Outcome of treatment in childhood acute lymphoblastic leukemia with rearrangements of the 11q23 chromosomal region. *Lancet* 2002;359:1909-1915.
18. Gleissner B, Goekbuget N, Rieder H, et al: CD10- pre-B acute lymphoblastic leukemia (ALL) is a distinct high-risk subgroup of adult ALL associated with a high frequency of MLL aberrations: results of the German Multicenter Trials for Adult ALL (GMALL). *Blood* 2005;106:4054-4056.
19. Kishi S, Cheng C, French D, et al: Ancestry and pharmacogenetics of antileukemic drug toxicity *Blood* 2007;109:4151-4157.
20. Yeoh EJ, Ross ME, Shurtleff SA, et al: Classification, subtype discovery, and prediction of outcome in pediatric acute lymphoblastic leukemia by gene expression profiling. *Cancer Cell* 2002;2:133-143.
21. Rocha JC, Cheng C, Liu W, et al: Pharmacogenetics of outcome in children with acute lymphoblastic leukemia. *Blood* 2005;105:4752-4758.
22. Cheok MH, Evans WE. Acute lymphoblastic leukemia: a model for the pharmacogenomics of cancer therapy. *Nat Rev Cancer* 2006;6:117-129.
23. Evans WE, Relling MV. Moving towards individualized medicine with pharmacogenomics. *Nature* 2004;429:464-468.
24. Holleman A, Cheok MH, den Boer ML, et al: Gene-Expression Patterns in Drug-Resistant Acute Lymphoblastic Leukemia Cells and Response to Treatment. *N Engl J Med* 2004;351:533-542.
25. Pui CH, Campana D, Evans WE. Childhood acute lymphoblastic leukemia – current status and future perspectives. *Lancet Oncol* 2001;2:597-607.
26. Simone J, Aur RJA, Hustu HO, et al : "Total Therapy" studies of acute lymphocytic leukemia in children: Current results and prospects for cure. *Cancer* 1972;30:1488-1494.
27. Holland JF, Glidewell O. Chemotherapy of acute lymphocytic leukemia of childhood. *Cancer* 1972; 30:1480-1487.
28. Ortega JA, Nesbit ME, Donaldson MH, et al: L-Asparaginase, vincristine, and prednisone for induction of first remission in acute lymphocytic leukemia. *Cancer Res* 1977;37:535-540.
29. Sallan SE, Cammita BM, Cassady JR, et al : Intermittent combination chemotherapy with adriamycin for childhood acute lymphoblastic leukemia: clinical results. *Blood* 1978;51:425-433.
30. Hitchcock-Bryan S, Gelber R, Cassady JR, et al: The impact of induction anthracycline on long-term failure-free survival in childhood acute lymphoblastic leukemia. *Med Pediatr Oncol* 1986;14:211-215.
31. Tubergen DG, Gilchrist GS, O'Brien RT, et al: Improved outcome with delayed intensification for children with acute lymphoblastic leukemia and intermediate presenting features: a Children's Cancer Group phase III trial. *J Clin Oncol* 1993;11:527-537.
32. Gaynon PS, Steinherz PG, Bleyer WA, et al : Improved therapy for children with acute lymphoblastic leukemia and unfavorable presenting features: A follow-up report of the Children's Cancer Group Study CCG-106. *J Clin Oncol* 1993;11:2234-2242.
33. Thomas DA, Faderl S, Cortes J, et al: Treatment of Philadelphia chromosome-positive acute lymphocytic leukemia with hyper-CVAD and imatinib mesylate. *Blood* 2004;103:4396-4407.
34. Yanada M, Takeuchi J, Sugiura I, et al: High complete remission rate and promising outcome by combination of imatinib and chemotherapy for newly diagnosed BCR-ABL-positive acute lymphoblastic leukemia: a phase II study by the Japan Adult Leukemia Study Group. *J Clin Oncol* 2006;24:460-466.
35. de Labarthe A, Rousselot P, Huguet-Rigal F, et al : Imatinib combined with induction or consolidation chemotherapy in patients with de novo Philadelphia chromosome-positive acute lymphoblastic leukemia: results of the GRAAPH-2003 study. *Blood* 2007;109:1408-1413.
36. Balis FM, Lester CM, Chrousos GP, et al: Differences in cerebrospinal fluid penetration of corticosteroids: possible relationship to the prevention of meningeal leukemia. *J Clin Oncol* 1987;5:202-207.
37. Jones B, Freeman AI, Shuster JJ, et al: Lower incidence of meningeal leukemia when prednisone is replaced by dexamethasone in the treatment of acute lymphocytic leukemia. *Med Pediatr Oncol* 1991;19:269-275.
38. Ito C, Evans WE, McNinch L, et al: Comparative cytotoxicity of dexamethasone and prednisolone in childhood acute lymphoblastic leukemia. *J Clin Oncol* 1996;14:2370-2376.
39. Kaspers GJ, Veerman AJ, Pop-Snijders C, et al: Comparison of the antileukemic activity in vitro of dexamethasone and prednisolone in childhood acute lymphoblastic leukemia. *Med Pediatr Oncol* 1996; 27:114-121.
40. Veerman AJ, Hahlen K, Kamps WA, et al: High cure rate with a moderately intensive treatment regimen in non-high-risk childhood acute lymphoblastic leukemia. Results of protocol ALL VI from the Dutch Childhood Leukemia Study Group. *J Clin Oncol* 1996;14:911-918.
41. Bostrom BC, Sensel MR, Sather HN, et al: Dexamethasone versus prednisone and daily oral versus weekly intravenous mercaptopurine for patients with standard-risk acute lymphoblastic leukemia: a report from the Children's Cancer Group. *Blood* 2003;101:3809-3817.
42. Mitchell CD, Richards SM, Kinsey SE, et al: Benefit of dexamethasone compared with prednisolone for childhood acute lymphoblastic leukaemia: results

- of the UK Medical Research Council ALL97 randomized trial. *Br J Haematol* 2005;129:734-745.
43. Igarashi S, Manabe A, Ohara A, et al: No advantage of dexamethasone over prednisolone for the outcome of standard- and intermediate-risk childhood acute lymphoblastic leukemia in the Tokyo Children's Cancer Study Group L95-14 protocol. *J Clin Oncol* 2005;23:6489-6498.
44. Hurwitz CA, Silverman LB, Schorin MA, et al: Substituting dexamethasone for prednisone complicates remission induction in children with acute lymphoblastic leukemia. *Cancer* 2000;88:1964-1969.
45. Pinheiro JP, Boos J. The best way to use asparaginase in childhood acute lymphoblastic leukemia – still to be defined? *Br J Haematol* 2004;125:117-127.
46. Wetzler M, Sanford BL, Kurtzberg J, et al: Effective asparagine depletion with pegylated asparaginase results in improved outcomes in adult acute lymphoblastic leukemia: Cancer and Leukemia Group B Study 9511. *Blood* 2007;109:4164-4167.
47. Nachman JB, Sather HN, Sensel MG, et al: Augmented Post-Induction Therapy for Children with High-Risk Acute Lymphoblastic Leukemia and a Slow Response to Initial Therapy. *NEJM* 1998;338: 1663-1671.
48. Pui CH, Simone JV, Hancock ML, et al: Impact of three methods of treatment intensification on acute lymphoblastic leukemia in children: long-term results of St Jude total therapy study X. *Leukemia* 1992;6:150-157.
49. Silverman LB, Gelber RD, Young ML, et al: Induction failure in acute lymphoblastic leukemia of childhood. *Cancer* 1999;85:1395-1404.
50. Appelbaum FR. Allogeneic hematopoietic stem cell transplantation for acute leukemia. *Semin Oncol* 1997;24:114-123.
51. Pui CH, Campana D. New definition of remission in childhood acute lymphoblastic leukemia. *Leukemia* 2000;14:783-785.
52. Szczepanski T, Orfao A, van der Velden VH, et al: Minimal residual disease in leukaemia patients. *Lancet Oncol.* 2001;2:409-417.
53. Sallan SE, Hitchcock-Bryan S, Gelber R, et al: Influence of intensive asparaginase in the treatment of childhood non-T-cell acute lymphoblastic leukemia. *Cancer Res* 1983;43:5601-5607.
54. Silverman LB, Gelber RD, Dalton VK, et al: Improved outcome for children with acute lymphoblastic leukemia: Results of Dana-Farber Consortium Protocol 91-01. *Blood* 2001;97:1211-1218.
55. Goldberg JM, Silverman LB, Levy DE, et al: Childhood T-cell acute lymphoblastic leukemia: The Dana-Farber Cancer Institute acute lymphoblastic leukemia consortium experience. *J Clin Oncol* 2003;21:3616-3622.
56. Camitta B, Leventhal B, Sauer S, et al: Intermediate-dose intravenous methotrexate and mercaptopurine therapy for non-T, non-B acute lymphocytic leukemia of childhood: A Pediatric Oncology Group study. *J Clin Oncol* 1989;7:1539-1544.
57. Camitta B, Mahoney D, Leventhal B, et al: Intensive intravenous methotrexate and mercaptopurine treatment of higher-risk non-T, non-B acute lymphocytic leukemia: A Pediatric Oncology Group study. *J Clin Oncol* 1994;12:1383-1389.
58. Land VJ, Shuster JJ, Crist WM, et al: Comparison of two schedules of intermediate-dose methotrexate and cytarabine consolidation therapy for childhood B-precursor cell acute lymphoblastic leukemia: A Pediatric Oncology Group study. *J Clin Oncol* 1994;12:1939-1945.
59. Hutchinson RJ, Gaynon PS, Sather H, et al: Intensification of Therapy for Children With Lower-Risk Acute Lymphoblastic Leukemia: Long-Term Follow-Up of Patients Treated on Children's Cancer Group Trial 1881. *J Clin Oncol* 2003;21:1790-1797.
60. Lange BJ, Bostrom BC, Cherlow JM, et al: Double-delayed intensification improves event-free survival for children with intermediate-risk acute lymphoblastic leukemia: a report from the Children's Cancer Group. *Blood* 2002;99:825-833.
61. Chessells M, Harrison G, Lillieyman JS, et al: Continuing (maintenance) therapy in lymphoblastic leukaemia: lessons from MRC UKALLX. *Br J Haematol* 1997;98:945-951.
62. Relling MV, Hancock ML, Boyett JM, et al: Prognostic Importance of 6-Mercaptopurine Dose Intensity in Acute Lymphoblastic Leukemia. *Blood* 1999;93:2817-2823.
63. Rivard GE, Hoyoux C, Infante C, et al: Maintenance chemotherapy for childhood acute lymphoblastic leukaemia: better in the evening. *Lancet* 1985;326:1264-1266.
64. Rivard GE, Lin KT, Leclerc JM, et al: Milk could decrease the bioavailability of 6-mercaptopurine. *Am J Pediatr Hematol Oncol* 1989;11:402-406.
65. Evans WE, Horner M, Chu YQ, et al: Altered mercaptopurine metabolism, toxic effects, and dosage requirement in a thiopurine methyltransferase-deficient child with acute lymphocytic leukemia. *J Pediatr* 1991;119: 985-989.
66. Lennard L, Gibson BE, Nicole T, et al: Congenital thiopurine methyltransferase deficiency and 6-mercaptopurine toxicity during treatment for acute lymphoblastic leukaemia. *Arch Dis Child* 1993;69: 577-579.
67. Relling MV, Hancock ML, Rivera GK, et al: Mercaptopurine therapy intolerance and heterozygosity at the thiopurine S-methyltransferase gene locus. *J Nat Cancer Inst* 1999;91:2001-2008.
68. Pui CH, Relling MV. Topoisomerase II inhibitor-related acute myeloid leukaemia. *Br J Haematol* 2000;109:13-23.
69. Relling MV, Rubnitz JE, Rivera GK, et al: High incidence of secondary brain tumours after radiotherapy and antimetabolites. *Lancet* 1999;354:34-39.
70. Thomsen JB, Schröder H, Kristinsson J, et al: Possible carcinogenic effect of 6-mercaptopurine on bone marrow stem cells, relation to thiopurine metabolism. *Cancer* 1999;86:1080-1086.
71. Harms DO, Göbel U, Spaar HJ, et al: Thioguanine offers no advantage over mercaptopurine in maintenance treatment of childhood ALL: results of the randomized trial COALL-92. *Blood* 2003;102: 2736-2740.
72. Matloub Y, Lindemulder S, Gaynon PS, et al: Intrathecal triple therapy decreases central nervous system relapse but fails to improve event-free survival when compared with intrathecal methotrexate: results of the Children's Cancer Group (CCG) 1952 study for standard-risk acute lymphoblastic leukemia, reported by the Children's Oncology Group. *Blood* 2006;108:1165-1173.
73. Vora A, Mitchell CD, Lennard L, et al: Toxicity and efficacy of 6-thioguanine versus 6-mercaptopurine in childhood lymphoblastic leukaemia: a randomized trial. *Lancet* 2006;368:1339-1348.
74. Conter V, Valsecchi MG, Silvestri D, et al: Pulses of vincristine and dexamethasone in addition to intensive chemotherapy for children with intermediate-risk acute lymphoblastic leukaemia: a multicentre randomised trial. *Lancet* 2007;369:123-131.
75. Toyoda Y, Manabe A, Tsuchida M, et al: Six months of maintenance chemotherapy after intensified treatment for acute lymphoblastic leukemia of childhood. *J Clin Oncol* 2000;18:1508-1516.
76. Shuster JJ, Wacker P, Pullen P, et al: Prognostic significance of sex in childhood B-precursor acute lymphoblastic leukemia: a Pediatric Oncology Group Study. *J Clin Oncol* 1998;16:2854-2863.
77. Pui CH, Boyett JM, Relling MV, et al: Sex Differences in Prognosis for Children With Acute Lymphoblastic Leukemia. *J Clin Oncol* 1999;17:818-824.
78. Nesbit, Jr ME, Sather HN, Robison LL, et al: Randomized study of 3 years versus 5 years of chemotherapy in childhood acute lymphoblastic leukemia. *J Clin Oncol* 1983;1:308-316.
79. Miller DR, Leikin SL, Albo VC, et al: Three versus five years of maintenance therapy are equivalent in childhood acute lymphoblastic leukemia: a report from the Children's Cancer Study Group. *J Clin Oncol* 1989;7:316-325.
80. Pinkel D: Five-year follow-up of Total Therapy" of childhood acute lymphocytic leukemia. *JAMA* 1971;216:648-652.
81. Evans AE, Gilbert ES, Zandstra R: The increasing incidence of central nervous system leukemia in children (Children's Cancer Study Group A). *Cancer* 1970;26:404-409.
82. Hustu HO, Aur RJA, Verzosa MS, et al: Prevention of central nervous system leukemia by irradiation. *Cancer* 1973;32:585-597.
83. Manera R, Ramirez I, Mullins J, Pinkel D: Pilot studies of species-specific chemotherapy of childhood acute lymphoblastic leukemia using genotype and immunophenotype. *Leukemia* 2000;8:1354-1361.
84. Pui CH, Howard S: Current management and challenges of malignant disease in the CNS in paediatric leukaemia. *Lancet Oncol* 2008;9:257-268.
85. Mahmoud HH, Rivera GK, Hancock ML, et al: Low leukocyte counts with blast cells in cerebrospinal fluid of children with newly diagnosed acute lymphoblastic leukemia. *N Engl J Med* 1993; 329:314-319.
86. Burger B, Zimmermann M, Mann G, et al: Diagnostic cerebrospinal fluid examination in children with acute lymphoblastic leukemia: Significance of low leukocyte counts with blasts or traumatic lumbar puncture. *J Clin Oncol* 2003;21:184-188.
87. Pui CH. Toward Optimal Central Nervous System-Directed Treatment in Childhood Acute Lymphoblastic Leukemia. *J Clin Oncol* 2003;21:179-181.

88. Tubergen DG, Gilchrist GS, O'Brien RT, et al: Prevention of CNS disease in intermediate-risk acute lymphoblastic leukemia: Comparison of cranial radiation and intrathecal methotrexate and the importance of systemic therapy: A Children's Cancer Group report. *J Clin Oncol* 1993;11:520-526.
89. Pullen J, Boyett J, Shuster J, et al: Extended triple intrathecal chemotherapy trial for prevention of CNS relapse in good-risk and poor-risk patients with B-progenitor acute lymphoblastic leukemia: A Pediatric Oncology Group study. *J Clin Oncol* 1993;11:839-849.
90. Conter V, Arico M, Valsecchi MG, et al: Extended intrathecal methotrexate may replace cranial irradiation for prevention of CNS relapse in children with intermediate-risk acute lymphoblastic leukemia treated with Berlin-Frankfurt-Munster-based intensive chemotherapy. *The Associazione Italiana di Ematologia ed Oncologia Pediatrica. J Clin Oncol* 1995;13:2497-2502.
91. Dibenedetto S, Di Cataldo A, Ragusa R, et al: Levels of L-asparagine in CSF after intramuscular administration of asparaginase from *Erwinia* in children with acute lymphoblastic leukemia. *J Clin Oncol* 1995;13:339-344.
92. Ribera J-M, Ortega J-J, Oriol A, et al: Comparison of Intensive Chemotherapy, Allogeneic, or Autologous Stem-Cell Transplantation As Postremission Treatment for Children With Very High Risk Acute Lymphoblastic Leukemia: PETHEMA ALL-93 Trial. *J Clin Oncol* 2007;25:16-24.
93. Balduzzi A, Valsecchi MG, Uderzo C, et al: Chemotherapy versus allogeneic transplantation for very-high-risk childhood acute lymphoblastic leukaemia in first complete remission: comparison by genetic randomization in an international prospective study. *Lancet* 2005;366:635-642.
94. Schrauder A, Reiter A, Gadner H, et al: Superiority of Allogeneic Hematopoietic Stem-Cell Transplantation Compared With Chemotherapy Alone in High-Risk Childhood T-Cell Acute Lymphoblastic Leukemia: Results From ALL-BFM 90 and 95. *J Clin Oncol* 2006;24:5742-5749.
95. Pieters R, Schrappe M, De Lorenzo P, et al: Outcome of infants less than one year of age with acute lymphoblastic leukemia treated with the Interfant-99 protocol. *Lancet*, in press.
96. Kosaka Y, Koh K, Kinukawa N, et al: Infant acute lymphoblastic leukemia with MLL gene rearrangements: outcome following intensive chemotherapy and hematopoietic stem cell transplantation. *Blood* 2004;104:3527-3534.
97. Sanders JE, Im HJ, Hoffmeister PA, et al: Allogeneic hematopoietic cell transplantation for infants with acute lymphoblastic leukemia. *Blood* 2005;105:3749-3756.
98. Sierra J, Radich J, Hansen JA, et al: Marrow transplants from unrelated donors for treatment of Philadelphia chromosome-positive acute lymphoblastic leukemia. *Blood* 1997;90:1410-1414.
99. Marks DI, Bird JM, Cornish JM, et al: Unrelated donor bone marrow transplantation for children and adolescents with Philadelphia-positive acute lymphoblastic Leukemia. *J Clin Oncol* 1998;16:931-936.
100. Biondi A, Cimino G, Pieters R, et al: Biological and therapeutic aspects of infant leukemia. *Blood* 2000;96:24-33.
101. Hilden JM, Dinndorf PA, Meerbaum SO, et al: Analysis of prognostic factors of acute lymphoblastic leukemia in infants: report on CCG 1953 from the Children's Oncology Group. *Blood* 2006;108: 441-451.
102. Thomas DA, O'Brien S, Jorgensen JL, et al: Prognostic significance of CD20 expression in adults with de novo precursor B-lineage acute lymphoblastic leukemia. *Blood* 2008 (prepub online August 14)
103. Rivera GK, Hudson MM, Liu Q, et al : Effectiveness of intensified rotational combination chemotherapy for late hematologic relapse of childhood acute lymphoblastic leukemia. *Blood* 1996; 88:831-837.
104. Gaynon PS, Qu RP, Chappell RJ, et al : Survival after relapse in childhood acute lymphoblastic leukemia : impact of site and time to first relapse—the Children's cancer Group experience. *Cancer* 1998;82:1387-1395.
105. Neale GA, Pui CH, Mahmoud HH, et al: Molecular evidence for minimal residual bone marrow disease in children with <isolated> extra-medullary relapse of T-cell acute lymphoblastic leukemia. *Leukemia* 1994;8:768-775.
106. Eckert C, Biondi A, Seeger K, et al: Prognostic value of minimal residual disease in relapsed childhood acute lymphoblastic leukaemia. *Lancet* 2001;358:1239-1241.
107. Coustan-Smith E, Gajjar A, Hijjiya N, et al: Clinical significance of minimal residual disease in childhood acute lymphoblastic leukemia after first relapse. *Leukemia* 2004;18:499-504.
108. Ritchey AK, Pollock BH, Lauer SJ, et al: Improved survival of children with isolated CNS relapse of acute lymphoblastic leukemia: A Pediatric Oncology Group Study. *J Clin Oncol* 1999;17:3745-3752.
109. Barredo JC, Devidas M, Lauer SJ, et al: Isolated CNS Relapse of Acute Lymphoblastic Leukemia Treated with Intensive Systemic Chemotherapy and Delayed CNS Radiation: A Pediatric Oncology Group Study. *J Clin Oncol* 2006;24:3142-3149.